(FILE 'HOME' ENTERED AT 07:06:32 ON 22 OCT 2001)

FILE 'BIOSIS, MEDLINE, INPADOC' ENTERED AT 07:08:41 ON 22 OCT 2001

L1 899 OUTER ROOT SHEATH

L2 26 L1 AND (CULTUR?/SA\FOLLICLE)

L3 17 DUPLICATE REMOVE L2 (9 DUPLICATES REMOVED)

=>

L Number	Hits	Search Text	DB	Time stamp
1	56	"outer root sheath"	USPAT; EPO; JPO; Derwent	2001/10/22 07:21

C:\APPS\EAST\workspaces\default.wsp

L3 ANSWER 14 OF 17 BIOSIS COPYRIGHT 2001 BIOSIS DUPLICATE 8
AN 1991:204710 BIOSIS
DN BA91:117405
TI GROWTH FACTORS SPECIFICALLY ALTER HAIR FOLLICLE CELL PROLIFERATION AND COLLAGENOLYTIC ACTIVITY ALONE OR IN COMBINATION.
AU WEINBERG W C; BROWN P D; STETLER-STEVENSON W G; YUSPA S H
CS LABORATORY CELLULAR CARCINOGENESIS TUMOR PROMOTION, DIVISION CANCER ETIOLOGY, NATIONAL CANCER INSTITUTE, BETHESDA, MD 20892, USA.
SO DIFFERENTIATION, (1990) 45 (3), 168-178.
CODEN: DFFNAW. ISSN: 0301-4681.
FS BA; OLD
LA English
AB A three-dimensional culture model for isolated murine pelage hair follicles in a type I collagen gel had been utilized to study the effects of selected growth factors on follicle cell proliferation and release of collagenolytic factors. **Cultured follicle** organoids differentially express cytokeratins 6 and 14 in a pattern suggesting they contain cells of the **outer root sheath**, inner root sheath and follicle matrix. Using incorporation of [3H]thymidine as a measure of proliferation, follicle organoids show a peak of DNA synthesis between day 1 and 5 of culture, depending on plating density, and then have a low rate of DNA synthesis. Thymidine incorporation is stimulated by transforming growth factor-alpha (TGF-.alpha.) in a dose-dependent response. Only peripheral cells presumably of the **outer root sheath**, incorporate thymidine in basal or stimulated conditions. TGF-.beta.1 and TGF-.beta.2 inhibit constitutive cell proliferation and oppose growth stimulation by TGF-.alpha.. Hair follicles lyse the collagen gel matrix when exposed to certain cytokines. Epidermal growth factor (EGF) and TGF-.alpha. stimulate gel lysis, but TGF-.beta.1, TGF-.beta.2 and cholera toxin do not. Other skin-derived cells, such as interfollicular epidermal cells, dermal fibroblasts, or combinations thereof, do not lyse gels in this culture model even when exposed to growth factors. Combinations of EGF or TGF-.alpha. with TGF-.beta.1 or TGF-.beta.2 are synergistic for collagenase release. These cytokines stimulate release of multiple species of matrix metalloproteinases, but the 92-kDa and 72 kDa type IV procollagenases and their activated derivatives predominate on zymograms. In cytokine-stimulated follicles, both peripheral and centrally located cells in the organoids express the 72-kDa type IV collagenase and a similar immunostaining pattern is present in developing follicles in vivo. Thus growth factors appear to work in concert for certain hair follicle responses and in opposition for others. These combined actions may play a role in different phases of hair follicle development that require cell replication and invasion into the deeper dermis.

L3 ANSWER 15 OF 17 BIOSIS COPYRIGHT 2001 BIOSIS
AN 1990:518249 BIOSIS
DN BA90:135525
TI ARCHITECTURE OF RECONSTRUCTED EPIDERMIS ON COLLAGEN LATTICES VARIES ACCORDING TO THE METHOD USED A COMPARATIVE STUDY.
AU LENOIR M C; BERNARD B A
CS CELL BIOL. DEP., CENT. INT. RECH. DERMATOLOGIQUES GALDERMA, F 06565 VALBONNE CEDEX, FR.
SO SKIN PHARMACOL, (1990) 3 (2), 97-106.

CODEN: SKPHEU.
FS BA; OLD
LA English
AB Epidermis was obtained in vitro after air exposure of keratinocyte cultures grown on a dermal equivalent. Some cultures were established from enzymatically dissociated keratinocytes of either interfollicular epidermis or hair follicle **outer root sheath**. Others resulted from centrifugal outgrowth of epidermal sheet, out of skin biopsies or hair follicles, which were directly implanted into dermal equivalents. Whatever the system used, a multilayered epidermis was obtained with an overall architecture resembling that of human epidermis. However, depending on the tissue culture method used and the source of keratinocytes, significant differences were observed. The most striking finding was the difference in 67 kDa keratin expression: the only case where it was strictly suprabasal and homogeneously expressed in the cytoplasm of the cells, as in normal epidermis, was found in the epidermis obtained from follicle explants. With the other methods, the expression of this marker was delayed and patchy. These results are discussed in term of possible intrinsic differences between interfollicular and follicular keratinocytes.

L3 ANSWER